Congenital Cytomegalovirus Infection in Twin Pregnancies: Viral Load in the Amniotic Fluid and Pregnancy Outcome

Tiziana Lazzarotto, PhD*; Liliana Gabrielli, MD*; Maria Pia Foscini, MD§; Marcello Lanari, MD‡; Brunella Guerra, MD§; Vincenzo Eusebi, MD¶; and Maria Paola Landini, MD*

ABSTRACT. Human cytomegalovirus (CMV) is the most common cause of viral intrauterine infection and fetal damage largely attributable to maternal primary infection. Most cases of congenital CMV infection in twins reported in the literature involved only 1 twin. We assessed the validity of polymerase chain reaction (PCR) and quantitative PCR on amniotic fluid (AF), at 21 to 22 weeks' gestation and at least 6 to 8 weeks after seroconversion, to predict the outcome of newborns in twin pregnancies. Two pregnant women with twin pregnancies and 1 woman with a triple pregnancy with primary CMV infection defined by the presence of immunoglobulin (Ig) M and low IgG avidity and/or by the presence of clinical symptoms and abnormal liver enzyme values were evaluated. CMV infection was found in 6 fetuses/newborns, 3 of whom were symptomatic. In the first twin pregnancy with diamniotic-dichorionic separate placentas, CMV symptomatic infection of the female twin was demonstrated by positive virus isolation and high viral load in AF. The male fetus was not infected as demonstrated by negative CMV culture and DNA detection in AF. In the triple pregnancy, the woman had a placenta with 2 monozygotic twins (females) and a separate placenta with a heterozygotic twin (male). The quantitative PCR results were 10^7 genome equivalents (GE)/mL of females AF and 1.9 × 10^6 GE/mL of male AF. Both female twins were asymptomatic at birth, whereas the male presented petechiae, thrombocytopenia, and cerebral ventriculomegaly. In the last twin pregnancy with fused dichorionic placentas, congenital CMV infection of both twins was diagnosed at birth in contrast with prenatal diagnosis. At time of amniocentesis, the left side twin was not infected as shown by negative results of CMV culture and DNA detection in AF. CMV infection of the right side twin was demonstrated by positive CMV DNA detection with a CMV DNA load of 4.9 × 10^4 GE/mL and positive virus isolation in the AF. The morphologic and histologic examinations of the placentas strongly supported a prenatal horizontal acquisition of CMV infection. These twin pregnancies showed a marked difference in the quantity of virus load documented by the prenatal diagnosis suggesting that twin fetuses may react differently to primary maternal infection despite being exposed to the same maternal influences. A high viral load is correlated with congenital CMV infections symptomatic at birth. In such cases, with fetal infection of only 1 twin (at amniocentesis) and fusion of placentas, fetal outcome of both twins needs to be evaluated for the possibility of viral transfer from one fetus to the other. Pediatrics 2003;112:e153–e157. URL: http://www.pediatrics.org/cgi/content/full/112/2/e153; cytomegalovirus, congenital infection, twin pregnancies, placenta, prenatal diagnosis, viral load.

ABBREVIATIONS. CMV, cytomegalovirus; PCR, polymerase chain reaction; AF, amniotic fluid; Ig, immunoglobulin; GE, genome equivalents; qPCR, quantitative polymerase chain reaction; ABR, auditory brainstem-evoked response; PMNL, polymorphonuclear leukocyte; EIA, enzyme immunoassay.

Cytomegalovirus (CMV) is the most common cause of intrauterine infection, affecting from 0.3% to 2% of liveborn infants. Between 10% and 15% of infants infected with congenital CMV exhibit the clinically apparent or symptomatic form of the disease, characterized by petechiae, hepatomegaly, splenomegaly, jaundice, periventricular calcifications, microcephaly, hearing impairment, and chorioretinitis. The remaining 85% to 90% of infected infants are asymptomatic at birth, but ~15% of them will develop delayed sequelae, especially progressive hearing loss.1

Primary infection in the mother and as intrauterine transmission during the first 16 weeks of pregnancy2 have a much greater clinical impact on the fetus than nonprimary infections and infections occurring during the last trimester of pregnancy.3–5

Intrauterine transmission occurs transplacentally during maternal viremia; the placenta acts as a portal of entry for the virus, but it also acts as a barrier because even during maternal primary infection, transmission occurs in only 40% of cases.4

Intrauterine transmission can be determined by both polymerase chain reaction (PCR) and virus isolation from amniotic fluid (AF).6–11 In single pregnancies, a high viral load in the AF seems to identify fetuses at higher risk of developing a severe infection, although the results should still be considered at an investigational stage because of the relatively low number of cases tested.12–14

A few cases of CMV-infected twin pregnancies are described in the literature, and often only 1 newborn was found infected.15,16 In only 1 case was prenatal diagnosis performed.17

Twin pregnancies represent an interesting model.
because different fetuses are simultaneously exposed to the same maternal influences. This paper describes 3 cases of CMV-infected twin pregnancies in which the maternal viral load in AF correlated with pregnancy outcome.

**CASE REPORTS**

**Case A**

A 23-year-old primiparous woman had a twin pregnancy (a female and a male fetus) that was without complication until 15 weeks’ gestation, when routine CMV serologic tests revealed suspected CMV infection (Table 1). Further investigations for CMV were performed. CMV-immunoglobulin (Ig) G with low avidity and CMV-IgM were demonstrated. Isolation of CMV from urine and saliva was negative, and viral antigenemia and DNAemia were negative. At gestation week 22, AF samples were collected from both sacs. CMV infection of the female twin was demonstrated by positive CMV DNA detection (3 × 10⁶ genome equivalents [GE]/mL of AF) and positive virus isolation. The male fetus was not infected as shown by negative CMV culture and DNA detection in the AF. At 31 weeks’ gestation, a discordant growth between the fetuses was diagnosed by ultrasound examination, revealing an intrauterine growth restriction in the female fetus. Pregnancy was complicated by in utero death of the female twin at gestation week 34. A cesarean section was performed 2 days later. The surviving infant had Apgar scores of 7 at 1 minute and 9 at 5 minutes and weighed 1700 g. At birth, no clinical evidence of congenital infection was detected, and the urine and saliva culture for CMV was negative. At autopsy, the female fetus showed hepatosplenomegaly and cerebral ventriculomegaly. Microscopic examination disclosed enlarged, inclusion-bearing cells in the lungs, liver, and kidneys. The overall features were characteristic of generalized cytomegalic inclusion disease.

Placentas were diagnostically dichorionic and separate. No histopathologic examination of placentas was done.

**Case B**

A 28-year-old woman had a triple pregnancy. She was a second gravida and had a placenta with 2 monzygotic twins (females) and a separate placenta with a heterozygotic twin (male; Table 1). At week 13 she had both anti-CMV-IgM and IgG with a low IgG avidity value, indicating a recent primary infection. Isolation of CMV from urine and saliva was negative, and the viral antigenemia and DNAemia were negative. AF was obtained at 21 weeks’ gestation from the fluid compartment of the heterozygotic fetus and from only 1 fluid compartment of the monzygotic twins. AF from the third gestational sac was not obtained for technical reasons. CMV DNA detection and virus isolation were positive in both AFs. The quantitative PCR (qPCR) results were: 10⁵ GE/mL of female AF and 1.9 × 10⁶ GE/mL of male AF. Delivery by cesarean section was planned at the 33rd week of pregnancy. The birth weights of the 2 female infants were 1350 and 1960 g, respectively. Apgar scores were 9 at 1 minute and 10 at 5 minutes. Both female twins were asymptomatic at birth. Ultrasound examination of the brain did not demonstrate significant abnormalities. Fundus oculi showed no signs of retinitis. Audiologic testing was performed only at 1 year by auditory brainstem-evoked response (ABR). No responses were noted at 100 dB SPL normal-hearing level in the right and left ears of both twins, indicating severe bilateral hearing loss.

The birth weight of the male infant was 1960 g; Apgar scores were 8/1 minute and 9/5 minutes. At physical examination the infant presented petechiae and laboratory assay showed thrombocytopenia. Ophthalmologic investigation was normal. Cranial ultrasonography showed ventriculomegaly. After a few days the infant’s physical status stabilized. Audiologic testing was performed only at 1 year. ABR were absent at 100 dB in the left ear, indicating monolateral sensorineural hearing loss.

**Case C**

A 24-year-old primipara with a twin pregnancy (2 female fetuses) had nausea, anorexia, fever, headache, and muscle weakness during the 15th week of pregnancy (Table 1). Biochemical tests detected raised blood transaminase levels; serologic tests disclosed both anti-CMV IgM and IgG, indicating a recent primary infection. Further, viral antigenemia (2pp65-positive-cells/2 × 10⁵ polymorphonuclear leukocytes [PMNLs]) and DNAemia (1.7 × 10⁵ GE/10⁵ PMNLs) were detected. Both urine and saliva were negative for CMV isolation. AF was collected only at the 2 gestational sacs by transabdominal amniocentesis at 21 weeks’ gestation. At the time of amniocentesis, maternal blood was CMV-negative. The left-side twin was not infected, as shown by negative CMV culture and DNA detection in the AF. CMV infection of the right-side twin was demonstrated by positive CMV DNA detection with a CMV DNA load of 4.9 × 10⁶ GE/mL and positive virus isolation in the AF. Routine ultrasonography examination showed normal growth of both fetuses. Cesarean delivery was performed at 36 weeks’ gestation. Both newborns were vigorous: Apgar scores at 1 and 5 minutes were 9 in both determinations for the left-side twin, the first born (weight 2670 g), and 7 and 9, respectively, for the right-side twin (weight 2400 g). In contrast with prenatal diagnosis, congenital infection of both newborns was diagnosed by a positive virus isolation in saliva and urine collected 1 day after birth. Although the left-side twin was antigenemia-negative and had a viral DNAemia of 7.8 × 10⁵ GE/10⁵ PMNLs, the right-side twin had 2 pp65-positive cells/2 × 10⁵ PMNLs and a DNAemia of 2.5 × 10⁵ GE/10⁵ PMNLs, suggesting an infection of the left-side twin after amniocentesis.

Further clinical studies revealed no cranial ultrasonography abnormalities and no evidence of chorioretinitis. However, ABR were absent at 80 dB in the right side twin, indicating severe hearing impairment caused by infection early in pregnancy. At 12 weeks of life,
months of life, the infant presented a monolateral sensorineural hearing loss.

The other infant had normal ABR and at 12 months of life was healthy, without auditory impairment.

Gross and histologic examination of the placenta showed a dichorionic, diamniotic-fused placenta. At submacroscopic examination, performed with a stereomicroscope and on histology, blood vessels crossing the 2 placentas were detected. Both placentas were CMV-positive, as detected by in situ hybridization.

METHODS

Serologic Tests

Anti-CMV IgG was evaluated with a commercial kit (Enzygnost anti-CMV/IgG enzyme immunoassay (EIA) α method; Behring, Marburg, Germany). Plates were read on a micro-EIA automatic reader (Behring). Anti-CMV IgM was evaluated using an anti-CMV/IgM kit (Enzygnost; Behring). Both kits were used and the results interpreted as suggested by the manufacturer. IgG avidity was determined using a commercial kit (Cytomegalovirus IgG Avidity EIA Well; Radim, Rome, Italy). The results were interpreted as suggested by the manufacturer; in particular, an avidity index (absorbance reading after urea wash/absorbance reading without urea wash × 100) <35% was considered low, >45% was considered high, and between 35% and 45% was considered moderate.

Virologic Tests

CMV Isolation

The shell vial procedure was used for CMV isolation from urine, saliva, and AF. The cells were fixed 24 to 48 hours after inoculation and were stained by an indirect immunofluorescence assay with a monoclonal antibody reacting with the CMV IE1 and E4 gene product (E13 + 2A2; Argene, Varilhes, France).

Antigenemia

The presence of CMV pp65 (ppUL83) in 2 × 10⁶ PMNLs of patients was determined, as recently described, using a CMV pp65-specific pool of 2 monoclonal antibodies (IC3 and AYM-1; Argene) in indirect immunofluorescence test.

PCR in AF

CMV DNA was individually extracted from 3 to 6 aliquots of AF (100 μL each) with an IsoQuick Nucleic Acid Extraction kit (Orca Research, Bothell, WA), and PCR was conducted as described in detail previously. AF was considered positive if at least 1 of the aliquots was positive.

PCR in Blood

CMV-DNA was extracted from aliquots of 1 × 10⁶ PMNLs by CMV-Ibridoquant Extraction kit (Bioline, Turin, Italy). Nested PCR was performed for amplification of a highly conserved region of the viral genome (major immediate-early) using 2 primer sets described elsewhere. PCR products were separated by electrophoresis on a 2% Nusieve-Seakem (2:1) gel (FMC Bioproducts) and visualized with transillumination. The sample was regarded as positive when a band corresponding to a 110-bp DNA fragment was detected. To exclude false positives owing to contamination, for each PCR batch, 4 samples containing the reaction mixture but no target DNA were processed. All samples containing no target DNA yielded negative results.

qPCR in Blood and in AF

Competitive qPCR was conducted with a commercially available kit (Amplification set CMV-Ibridoquant, Bioline). DNA was extracted from aliquots of 1 × 10⁶ PMNLs by CMV-Ibridoquant Extraction kit (Bioline) and from AF as previously described. PCR was run as suggested by the manufacturers. In this test, an internal control construct, which is amplified by the same primer pairs but recognized by a different molecular probe, is amplified and detected. This method allows reproducible amplification of a minimum of 100 GE/1 × 10⁶ PMNLs and of 940 GE/mL of AF. Plasmids were used as positive controls.

The quantitative determination of CMV DNA in the AF of at least 10³ GE gave a 100% certainty of detecting an infected fetus/newborn. Higher viral loads (≥10⁵) were associated with fetuses or newborns with symptoms. To estimate the interassay variability of the qPCR assay, DNA extracted from the AF was quantified in 3 independent PCR runs. The mean GE value ± the standard deviation was 1827 ± 187 GE, and the coefficient of variation was 10.2%.

Morphologic and Histologic Examinations

The placenta was formalin fixed and then sectioned with a large knife perpendicular to fetal membranes.

For 3-dimensional tissue reconstruction, large sections (macrosections) were obtained including the insertions of the 2 umbilical cords and the membrane separating the 2 fetal cavities. Macrosections were fixed flat overnight, washed in tap water, dehydrated through a graded series of alcohol to xylol, and finally cleared in methylsalicylate. Macrosections were examined under a stereomicroscope (Nikon, Tokyo, Japan) and subsequently were paraffin-embedded and prepared for histologic examination.

For conventional histologic examination, small blocks of placental tissue were obtained and paraffin was embedded with a routine procedure. Blocks were cut and histologic sections stained with hematoxylin and eosin. From selected blocks, 4-μm sections were cut and pretreated for in situ hybridization as previously described. For in situ hybridization, a biotinylated DNA CMV probe was employed (Enzo Diagnostics Inc, Farmingdale, NY). The probe was denatured at 95°C for 10 minutes. Hybridization was performed overnight at 37°C in a humid chamber. The probe was diluted 1/10 in hybridization buffer supplied by Enzo Diagnostics. To enhance positivity, 2 cycles of catalyzed amplification system (Dako GenPoint; Dako) were performed. Negative controls consisted in omitting the probe; sections of known positivity were added as positive controls.

DISCUSSION

Maternal factors are thought to explain the variable outcome of congenital CMV infection in newborns after primary CMV infection during pregnancy.

Some have suggested that women who transmit CMV to the fetuses might have defective immunologic responses and, therefore, are unable to limit replication of the virus. In contrast, the results of our study are in agreement with literature reports showing that twin fetuses may react differently to primary maternal CMV infection, although exposed to the same maternal influences. Therefore, the defective immunologic response in the mother cannot alone account for the different outcome of transmission.

Congenital CMV infection is almost invariably accompanied by positive histopathologic findings in the placenta, indicating that the placenta is the main entrance for CMV to the fetuses and that placental infection following maternal viremia is the initiating step of fetal infection. The placenta could have a more important role as a protective factor than maternal immunologic reactivity.

One of the factors in limiting viral spread could be the ability of the placenta to produce interferon when challenged. Interferons are constitutively expressed by several tissues, notably in the placenta of several species including humans. Fisher et al observed that CMV infection impairs cytrophoblast expression of HLA-G, an important component of the mechanism that protects fetal cells from removal by maternal immune cells.

Another theory is that the placenta could act as a nonspecific barrier. In our dizygotic twins (case A) with separate pla-
CONGENITAL CYTOMEGALOVIRUS INFECTION IN TWINS

This study shows that maternal factors play a very limited role in influencing CMV transmission, as fetuses simultaneously exposed to the same maternal influence had a completely different outcome. Therefore, even sophisticated studies in the mother will not be sufficient to predict congenital infection in the fetus, and studies in the fetus are a necessary complement. In case of serologic diagnosis of maternal primary infection in early pregnancy, prenatal diagnosis should be offered to detect or exclude fetal infection and a high viral load predicts symptomatic congenital infection. Furthermore, in twin pregnancies with fetal infection of only 1 twin (at amniocentesis) and with fusion of placentas, prenatal diagnosis has limited value and the outcome of both twins needs to be evaluated for the possibility of a prenatal horizontal acquisition of the infection.

ACKNOWLEDGMENTS

This study was supported in part by grants from the Ministry of Public Health (Istituto Superiore di Sanità, AIDS Program); the Ministry of University, Scientific and Technological Research; the St. Orsola Malpighi General Hospital; and the University of Bologna.

We thank Cristiana Grandi and Elisabetta Magrini for excellent technical assistance. Anne Collins edited the English text.

REFERENCES


21. Tot T, Tabor I, Dean PB. The pressing need for better histologic-mammographic correlation of the many variations in normal breast anatomy. *Virchows Arch. 2000;437:338–344*


Congenital Cytomegalovirus Infection in Twin Pregnancies: Viral Load in the Amniotic Fluid and Pregnancy Outcome
Tiziana Lazzarotto, Liliana Gabrielli, Maria Pia Foschini, Marcello Lanari, Brunella Guerra, Vincenzo Eusebi and Maria Paola Landini

Pediatrics 2003;112:e153

Updated Information & Services
including high resolution figures, can be found at:
/content/112/2/e153.full.html

References
This article cites 35 articles, 7 of which can be accessed free at:
/content/112/2/e153.full.html#ref-list-1

Citations
This article has been cited by 3 HighWire-hosted articles:
/content/112/2/e153.full.html#related-urls

Subspecialty Collections
This article, along with others on similar topics, appears in the following collection(s):
Infectious Disease
/cgi/collection/infectious_diseases_sub

Permissions & Licensing
Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at:
/site/misc/Permissions.xhtml

Reprints
Information about ordering reprints can be found online:
/site/misc/reprints.xhtml
Congenital Cytomegalovirus Infection in Twin Pregnancies: Viral Load in the Amniotic Fluid and Pregnancy Outcome
Tiziana Lazzarotto, Liliana Gabrielli, Maria Pia Foschini, Marcello Lanari, Brunella Guerra, Vincenzo Eusebi and Maria Paola Landini

Pediatrics 2003;112;e153

The online version of this article, along with updated information and services, is located on the World Wide Web at:
/content/112/2/e153.full.html