Blood Cultures in the Evaluation of Children With Cellulitis

Karin Berger Sadow, MD*, and James M. Chamberlain, MD*‡

ABSTRACT. Objective. To evaluate the yield of blood cultures obtained from immunocompetent children admitted for cellulitis in the post-Haemophilus influenzae type b (Hib) vaccine era and to determine whether these cultures are cost-effective.

Design. Retrospective case series.

Setting. Urban pediatric emergency department.

Study Population. Patients 2 days to 22 years of age admitted with cellulitis from 1994 through 1995.

Measurements and Results. Of 381 patients identified, 266 (70%) had blood cultures and 243 of these children were enrolled. Data recorded include demographics, immunization status, initial clinical appearance, antibiotic pretreatment, preexisting illness, location and precipitating cause of cellulitis, white blood cell count, and band-to-neutrophil ratio (BNR). Blood cultures were categorized as positive, negative, or contaminant. Five cultures (26%) were positive, and 13 (5.4%) were contaminants. The positive blood cultures grew streptococcus and staphylococcus organisms, and none of the children were bacteremic with H influenzae. All patients with group A β-hemolytic streptococci had active varicella. The mean age was lower (26 vs 75 months) in those with a positive blood culture, and mean BNR was higher (0.32 vs 0.07). Patient management did not change for bacteremic patients with uncomplicated cellulitis. All repeat cultures were negative. The cumulative charge for all blood cultures was $50 986.

Conclusions. Blood cultures are not cost-effective and are more frequently contaminated than positive in the evaluation of a patient with uncomplicated cellulitis. Since introduction of the H influenzae type b vaccine, the most common organisms are streptococci. Using a BNR = 0.20 as a threshold for sending blood cultures, we would have missed one positive culture, but would have avoided blood cultures in 213 patients (88%) with an estimated savings of $42 850. Pediatrics 1998;101(3). URL: http://www.pediatrics.org/cgi/content/full/101/3/e4; cellulitis, bacteremia, health care costs.

ABBREVIATIONS. Hib, Haemophilus influenzae type b; ED, emergency department; WBC, white blood cell; BNR, band-to-neutrophil ratio; GABHS, group A β-hemolytic streptococcus; CI, confidence interval.

From the Departments of *Pediatrics and ‡Emergency Medicine, George Washington University School of Medicine and Health Sciences and Children’s Hospital, Washington, DC.

This work was presented in part at the Annual Meeting of the Ambulatory Pediatric Association, May 4, 1997, Washington, DC.

Received for publication Jul 21, 1997; accepted Nov 17, 1997.

Reprint requests to (K.B.S.) Children’s National Medical Center, 111 Michigan Ave, NW, Suite 1450, Washington, DC 20010.

PEDIATRICS (ISSN 0031 4005). Copyright © 1998 by the American Academy of Pediatrics.
Analysis

Statistical analysis was performed using Kwikstat and Epistat statistical software. Descriptive statistics are reported as means (±SD) and medians with ranges. The association of categorical variables with bacteremia was tested using the χ² test, and the association of continuous variables with bacteremia was tested with the t test and analysis of variance. A critical α of 0.05 was considered statistically significant.

RESULTS

During the 2-year study period, 381 inpatients were identified with a hospital discharge diagnosis of cellulitis. In 266 (70%) of these patients, blood cultures were obtained. Twenty-three patients were excluded (19 charts were unavailable, and 4 patients developed cellulitis after hospitalization). The remaining 243 comprised the study sample. During the same period, there were 108 patients with other infections having International Classification of Diseases, 9th ed, codes other than cellulitis who were not included in our study (4 patients with erysipelas, 22 with impetigo, and 82 with chickenpox with other complication).

The mean age of patients was 6.2 ± 5.0 years, and 57% were female. Sixty-seven percent were African-American, 16% white, and 11% Hispanic. Immunizations were current for 94% of patients, and only 22% were pretreated with antibiotics before admission (47 oral, 5 intramuscular, and 1 intravenous). The majority (91%) of patients had either no or noncontributory underlying disease. None of the patients with positive blood cultures were pretreated or had an underlying medical condition.

A total of 236 patients (97%) were described as nontoxic, 1 was described as toxic, and 1 was in shock. The toxic-appearing patient had cellulitis after pin removal for slipped capital femoral epiphysis, and the second patient was a hypotensive 15-year-old with a trunk cellulitis of unknown etiology. Both of these patients had negative blood cultures, although 1 was pretreated with intramuscular cefazolin.

The location and precipitating cause of cellulitis are depicted in Figs 1 and 2. Thirty percent of patients had cellulitis involving an extremity. Most cases of cellulitis (72%) were uncomplicated, but 60 (25%) of the patients had an abscess, 3 (1.2%) had osteomyelitis, 1 had a septic joint and osteomyelitis, 1 had a septic joint and psoas abscess, 1 had mastoiditis, 1 had tenosynovitis, and 1 had a thigh abscess and compartment syndrome.

Blood cultures were positive in 5 patients (2.1%; 95% confidence interval [CI]: 0.67, 4.74) and contaminated in 13 (5.4%; 95% CI: 2.88, 8.97). The positive blood cultures grew streptococcus (n = 4) and staphylococcus organisms (n = 1). The characteristics of these bacteremic patients are shown in Table 1. All patients bacteremic with GABHS had active infection with varicella. The patient with S aureus had a cellulitis complicated by a septic elbow and osteomyelitis, and the patient with S pneumoniae had a septic hip and psoas abscess. There were no positive blood cultures in the 53 pretreated patients (0%; 95% CI: 0, 6.7) and 5 positive blood cultures in 290 patients who were not pretreated (2.6%; 95% CI: 0.86, 6.0).

The mean age was lower (26 vs 75 months; P = .01) in those with a positive blood culture, and the mean BNR was higher (0.32 vs 0.07; P < .001). The mean temperature was 39.3°C in bacteremic patients and 38.0°C in nonbacteremic patients (P = .018). The mean WBC count for bacteremic patients was 23 600 ± 19 950/µL, and for nonbacteremic patients 14 870 ± 14 850/µL (P = .386).

For those patients with a positive culture and uncomplicated cellulitis, patient management did not change other than obtaining a repeat culture for every positive and contaminated culture. All repeat cultures were negative. The only change in antibiotic therapy was for a patient with a septic hip and psoas abscess; vancomycin was added to Fig 1. Location of cellulitis.

Fig 2. Precipitating cause of cellulitis.
the initial therapy of oxacillin and cefotaxime after the blood and abscess cultures grew *S. pneumoniae*.

The length of stay for the bacteremic patients was 14.6 ± 9.8 days, and for the nonbacteremic patients, 4.5 ± 4.2 days (P < .001). For the patients with uncomplicated cellulitis and positive blood cultures (*n = 2*), the lengths of stay were 3 and 11 days, similar to the mean length of stay in the negative/contaminated group. For the three bacteremic patients with associated diseases (one abscess and compartment syndrome, one osteomyelitis and septic arthritis, and one septic arthritis and psoas abscess), the lengths of stay were 19, 11, and 29 days.

The charge to patients associated with negative blood cultures was $44,775 ($128 per aerobic culture + $71 per anaerobic culture × 225 cultures). The charge for positive blood cultures was $1320 ($128 + $71 + $65 for sensitivities) × 5) and contaminated cultures $2587 ($128 + $71) × 13). The charge for repeat cultures was $2304 ($128 × 18). The cumulative charge associated with obtaining blood cultures was $50,986.

**DISCUSSION**

This study is the first to define the yield of blood cultures in children admitted for all types of cellulitis in the post-Hib vaccine era. It is also the first study to attempt to determine the cost-effectiveness of routine blood cultures in the management of these patients. Many previous studies and standard textbooks advocate obtaining blood cultures in the evaluation of patients with cellulitis, but this recommendation was based historically on the high frequency of bacteremia with *H. influenzae*.

In only 5 (2%) of 243 patients did the blood culture yield a true pathogen. Four of the five isolated pathogens were of the streptococcal species. This is consistent with the observations of Schwartz et al in their retrospective series of periorbital cellulitis in the post-Hib vaccine era. In our study, no children were bacteremic with *H. influenzae*.

Children with a true-positive blood culture result had a lower mean age and a higher mean BNR. Based on our data, we would have used a BNR = 0.20 as a threshold for sending blood cultures, we would have missed one positive blood culture (a 5-year-old with active varicella and GABHS bacteremia, BNR = 0.02), but would have avoided blood cultures in 213 patients (88%). This corresponds to an estimated savings of $42,850.

The association of GABHS with varicella in this study is of interest and has been observed in previous studies. Fifteen percent of the patients in our study with varicella and cellulitis were bacteremic, and all presented on the third to fifth day of illness. As described by Doctor et al, invasive GABHS infections must be considered in children with varicella and fever on or beyond the third day of illness. In their study, it was noted that the average WBC count of patients with varicella and bacteremia was not elevated (11,500 ± 8400/µL). Two of our patients with a positive blood culture and varicella had normal WBC counts (6000 and 12,000/µL, respectively), whereas the other had an elevated WBC count of 25,000/µL. The results of both studies suggest that a normal WBC count does not preclude the possibility of bacteremia with GABHS during varicella infection.

The patients bacteremic with *S. aureus* and *S. pneumoniae* both had complicated cellulitis; one had a septic elbow and osteomyelitis (*S. aureus*), and the other, a septic hip and psoas abscess. Both of these patients were <6 months of age. Because of their age and associated pathology, a blood culture was indicated as part of their complete evaluation.

Our study had several limitations. First, it is a retrospective study, thus, reports of clinical appearance and documentation of precipitating cause are limited to the patient record. Second, we may have missed some cases of bacteremia because 22% of the patients were pretreated with antibiotics before a blood culture was obtained. Finally, 30% of patients admitted did not have blood cultures performed.

Although the study design does not allow us to calculate the true incidence of bacteremia, several recommendations can be made based on this large case series. First, positive blood cultures are uncommon and were seen only in association with active varicella or underlying soft-tissue infections (osteomyelitis, septic arthritis). Second, blood cultures were twice as likely to be contaminated than positive. Third, obtaining blood culture studies involves significant costs, including financial and emotional costs (pain, etc) of repeating studies in the 5% of patients with contaminants. Finally, the small number of bacteremic patients precludes definitive statements about the prospective use of the complete blood count or BNR to predict bacteremia.

Most children with cellulitis are treated as outpatients. For children ill enough to be admitted, the associations noted with young age, high BNR, and varicella lend support to a practice of performing blood cultures in young children with any of these risk factors. However, another reasonable interpretation of our data would be to not obtain a complete blood count or blood culture unless the patient had varicella, an associated osteomyelitis or septic arthritis, or a toxic appearance.

In summary, blood cultures are not cost-effective

---

**TABLE 1.** Characteristics of Bacteremic Patients at Presentation

<table>
<thead>
<tr>
<th>Age (Months)</th>
<th>Organism</th>
<th>Height of Fever (°C)</th>
<th>BNR</th>
<th>Location</th>
<th>Associated Pathology</th>
</tr>
</thead>
<tbody>
<tr>
<td>60</td>
<td>GABHS</td>
<td>39.3</td>
<td>0.02</td>
<td>Extremity</td>
<td>Varicella</td>
</tr>
<tr>
<td>48</td>
<td>GABHS</td>
<td>39</td>
<td>0.28</td>
<td>Extremity</td>
<td>Varicella, abscess, and compartment syndrome</td>
</tr>
<tr>
<td>12</td>
<td>GABHS</td>
<td>40.5</td>
<td>0.58</td>
<td>Trunk</td>
<td>Varicella</td>
</tr>
<tr>
<td>4</td>
<td><em>Streptococcus pneumonia</em></td>
<td>39.4</td>
<td>0.40</td>
<td>Extremity</td>
<td>Septic hip and psoas abscess</td>
</tr>
<tr>
<td>6</td>
<td><em>Staphylococcus aureus</em></td>
<td>38.5</td>
<td>0.30</td>
<td>Extremity</td>
<td>Septic elbow and osteomyelitis</td>
</tr>
</tbody>
</table>

---

http://www.pediatrics.org/cgi/content/full/101/3/e4

Downloaded from http://pediatrics.aappublications.org/ by guest on November 13, 2017
in the management of immunocompetent patients admitted for cellulitis. The costs associated with obtaining blood cultures are substantial, and the results do not alter therapy. In addition, iatrogenic injury may result from repeating blood cultures when initial cultures are positive or contaminated. Clinicians who wish to obtain blood cultures should limit cultures to toxic-appearing children, patients with varicella, or those with another focus of infection, such as a septic joint.

ACKNOWLEDGMENTS

We thank Drs Bruce Klein and Douglas Boenning for reviewing this manuscript.

REFERENCES

Blood Cultures in the Evaluation of Children With Cellulitis
Karin Berger Sadow and James M. Chamberlain
Pediatrics 1998;101;e4
DOI: 10.1542/peds.101.3.e4

Updated Information & Services
including high resolution figures, can be found at:
http://pediatrics.aappublications.org/content/101/3/e4

References
This article cites 6 articles, 1 of which you can access for free at:
http://pediatrics.aappublications.org/content/101/3/e4.full#ref-list-1

Subspecialty Collections
This article, along with others on similar topics, appears in the following collection(s):
Hematology/Oncology
http://classic.pediatrics.aappublications.org/cgi/collection/hematology:oncology_sub
Infectious Disease
http://classic.pediatrics.aappublications.org/cgi/collection/infectious_diseases_sub

Permissions & Licensing
Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at:
https://shop.aap.org/licensing-permissions/

Reprints
Information about ordering reprints can be found online:
http://classic.pediatrics.aappublications.org/content/reprints

Pediatrics is the official journal of the American Academy of Pediatrics. A monthly publication, it has been published continuously since . Pediatrics is owned, published, and trademarked by the American Academy of Pediatrics, 141 Northwest Point Boulevard, Elk Grove Village, Illinois, 60007. Copyright © 1998 by the American Academy of Pediatrics. All rights reserved. Print ISSN: .